

Supplementary Information

Preparation of Magnetoliposomes with a Green, Low-Cost, Fast and Scalable Methodology and Activity Study against *S. aureus* and *C. freundii* Bacterial Strains

Rosângela M. F. da Costa e Silva,^{*a} Luciano R. S. Lara,^a Jorge L. López,^b
 Ângela L. Andrade,^c Junnia A. C. Oliveira,^d Jacqueline A. Takahashi,^a
 Henriete S. Vieira,^a Tulio Matencio,^a Humberto O. Stumpf^a and Rosana Z. Domingues^a

^aDepartamento de Química, Universidade Federal de Minas Gerais (UFMG),
 Av. Antonio Carlos 6627 Pampulha, 31270-901 Belo Horizonte-MG, Brazil

^bCentro de Ciências Biológicas e da Natureza (CCBN), Universidade Federal do Acre (UFAC),
 Rodovia BR 364, km 04, s/n, Distrito Industrial, 69915-900 Rio Branco-AC, Brazil

^cDepartamento de Química, Universidade Federal de Ouro Preto (UFOP), Morro do Cruzeiro,
 35400-000 Ouro Preto-MG, Brazil

^dInstituto de Ciências Biológicas, Universidade Federal de Minas Gerais (UFMG),
 Av. Antonio Carlos 6627 Pampulha, 31270-901 Belo Horizonte-MG, Brazil

Characterization of Fe₃O₄@lecithin

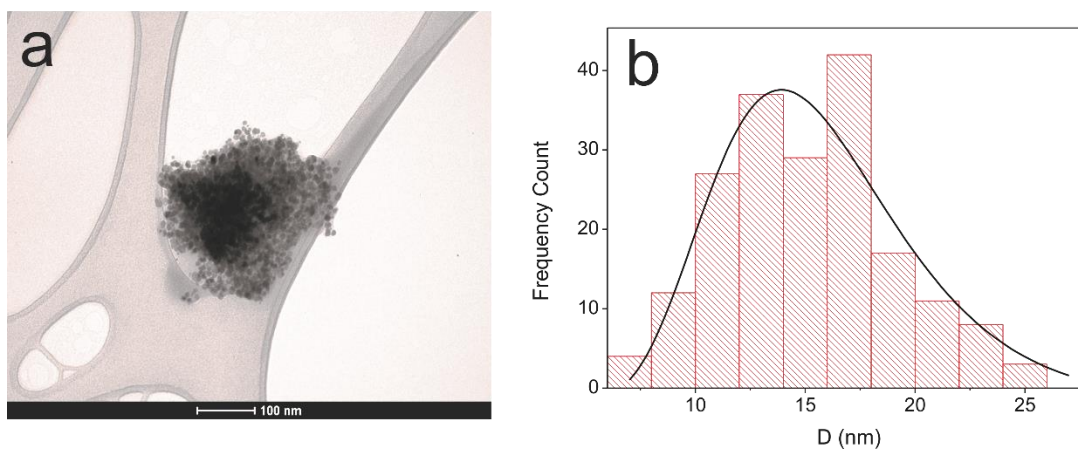


Figure S1. (a) Representative TEM of Fe₃O₄@lecithin; (b) distribution of size NPs of Fe₃O₄@lecithin.

The Langevin equation S1:

$$M = \int_0^{\infty} Ms \left[\left(\coth \left(\frac{\mu(D)H}{k_B T} \right) - \left(\frac{k_B T}{\mu(D)H} \right) \right) \frac{1}{\langle D \rangle \sigma_m \sqrt{2\pi}} \exp \left(- \frac{(\ln D / \langle D \rangle)^2}{2\sigma_m^2} \right) \right] dD + \chi H \quad (S1)$$

where $\mu(D)$ is the momentum of the particle as a function of diameter, k_B is the Boltzmann constant, M_s is the saturation magnetization, σ_m is the standard deviation of the particle size distribution, χ is the linear susceptibility term, H is the magnetic field intensity, T is the temperature, D is the diameter.

*e-mail: rosangela_ferreirafeliz@yahoo.com.br

Characterization of liposomes

The magnetite contents encapsulated in the Lip-Fe₃O₄@lecithin and Lip-Stigma-Fe₃O₄@lecithin samples were performed by AAS. The encapsulation of the magnetite was equal to 70% in Lip-Fe₃O₄@lecithin and 97% for Lip-Stigma-Fe₃O₄@lecithin described in Table 1 (main text). Therefore, the content of NPs encapsulated in Lip-Stigma-Fe₃O₄@lecithin was 27% higher in relation to Lip-Fe₃O₄@lecithin, attributed to the presence of stigmasterol. The variations of the encapsulated contents of magnetite were less than 5% during the 14-day period, confirmed by AAS. The liposomes show high encapsulation capacity of NPs. Hüsich *et al.*¹ describe systems with elevated encapsulated active content such as phospholipid complexes.

¹H NMR of Lip, stigmasterol and Lip-Stigma

The signal at δ 3.29 ppm (Figure S2a) was attributed to the presence of phosphatidylcholine (PC, -N(CH₃)₃) and δ 3.19 (-CH₂-NH₃⁺) and 8.26 ppm (NH₃⁺) were attributed to phosphatidylethanolamine (PE).¹⁻⁶ The signal at δ 5.22 ppm was attributed to the presence of the esterified glycerol of phospholipids (Gly, CH-OCOR).^{1,7} The presence of steroids (S) in Lip and lecithin was confirmed by the signal at δ 0.68 and 1.84 ppm.⁸⁻¹⁰ The determination of the molar ratio between the components was done using the following signals: δ 5.22 (Gly) and 0.68 ppm (S). Calculations were according to the literature.^{2,4,5}

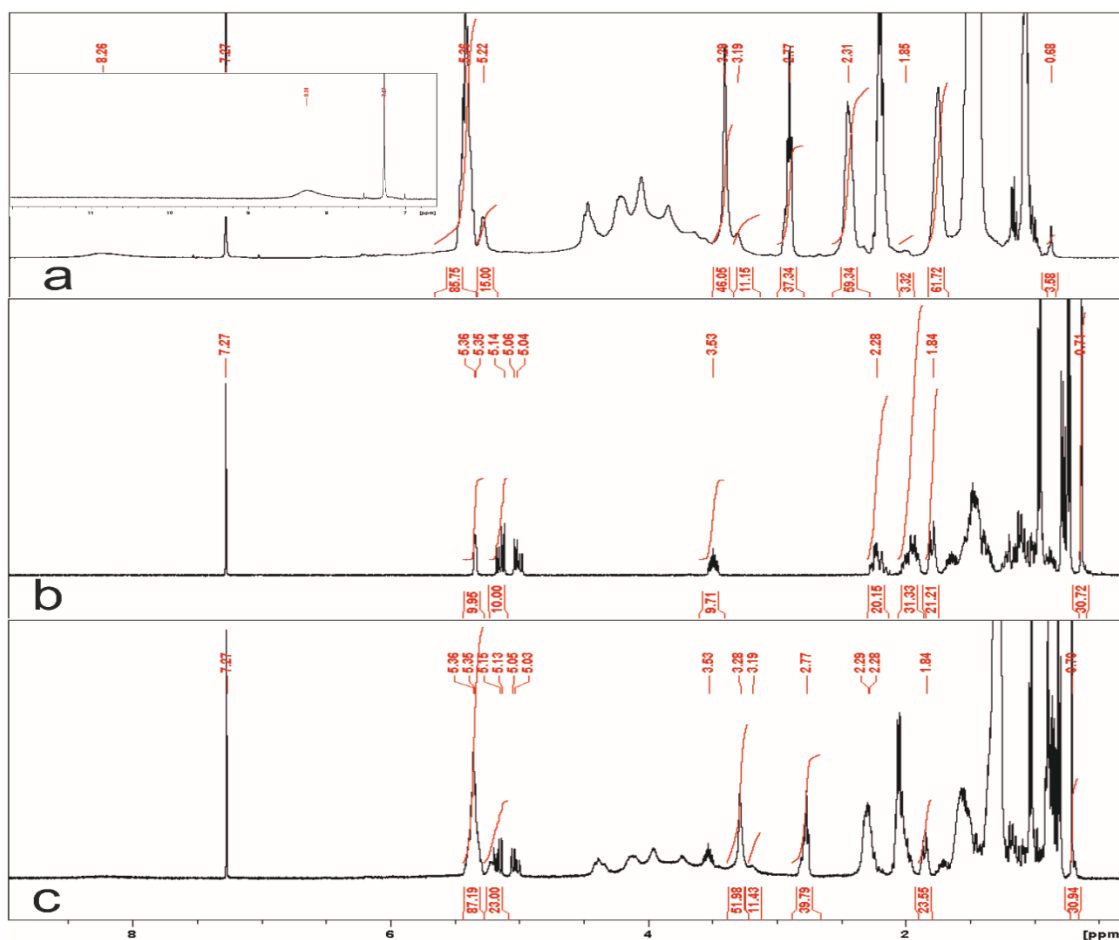


Figure S2. ¹H NMR spectra (400 MHz, CDCl₃) of Lip after 14 days stored in 4 °C (a); stigmasterol (b) and Lip-Stigma after 14 days stored in 4 °C (c).

The SAR of samples were calculate using the graphic of Figure S3a.

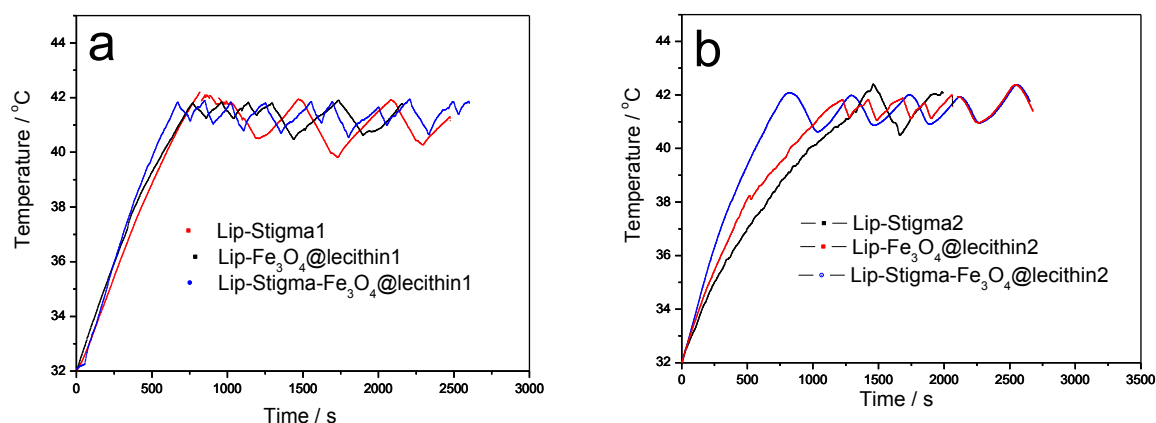


Figure S3. Heat rates of Lip-Stigma1, Lip-Fe₃O₄@lecithin1 and Lip-Stigma-Fe₃O₄@lecithin1 (a); and Lip-Stigma2, Lip-Fe₃O₄@lecithin2 and Lip-Stigma-Fe₃O₄@lecithin2 (b) in the presence of magnetic field (MF41) of 200 Oe, 315 kHz.

References

1. Hüsch, J.; Dutagaci, B.; Glaubitz, C.; Geppert, T.; Schneider, G.; Harms, M.; Müller-Goymann, C.; Fink, L.; Schmidt, M. U.; Setzer, C.; *Eur. J. Pharm. Sci.* **2011**, *44*, 103.
2. Mertins, O.; Sebben, M.; Schneider, P. H.; Pohlmann, A. R.; Silveira, N. P.; *Quim. Nova* **2008**, *31*, 1856.
3. Scholfield, C.; *J. Am. Oil Chem. Soc.* **1981**, *58*, 889.
4. Hatzakis, E.; Koidis, A.; Boskou, D.; Dais, P.; *J. Agric. Food Chem.* **2008**, *56*, 6232.
5. Troutier, A.-L.; Véron, L.; Delair, T.; Pichot, C.; Ladaviere, C.; *Langmuir* **2005**, *21*, 9901.
6. Cheung, A. P.; Olson, L. L.; *J. Pharm. Biomed. Anal.* **1990**, *8*, 729.
7. Vlahov, G.; *Prog. Nucl. Magn. Reson. Spectrosc.* **1999**, *35*, 341.
8. Clausen, M. R.; Edelenbos, M.; Bertram, H. C.; *J. Agric. Food Chem.* **2014**, *62*, 4392.
9. Li, W.-J.; Lin, Y.-C.; Wu, P.-F.; Wen, Z.-H.; Liu, P.-L.; Chen, C.-Y.; Wang, H.-M.; *Int. J. Mol. Sci.* **2013**, *14*, 1698.
10. Adosraku, R.; Choi, G.; Constantinou-Kokotos, V.; Anderson, M.; Gibbons, W.; *J. Lipid Res.* **1994**, *35*, 1925.