

Botocudorol, a New Bisabolene Type *nor*-Sesquiterpenoid from *Sparattanthelium botocudorum*

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Do extrato etanólico-acquoso (95%) das folhas de *Sparattanthelium botocudorum* foram isolados e identificados 5-hidroxi-3,4',7-trimetoxiflavona(1) e 4 α ,10 α -dihidroiguaja-6-eno (2) e um novo *nor*-sesquiterpeno, que foi denominado botocudorol (3). Dados espectrais foram usados nas determinações estruturais. As atribuições dos deslocamentos químicos dos átomos de ^1H e ^{13}C apoiaram-se na análise dos resultados fornecidos por experiências APT, DEPT, ^1H x ^1H -HOMOCOSY e ^1H x ^{13}C -HETECOSY ($^1\text{J}_{\text{CH}}$).

From the 95% ethanolic extract of the leaves of *Sparattanthelium botocudorum* were isolated and identified 5-hidroxy-3,4',7-trimethoxyflavone(1), 4 α ,10 α -dihydroiguaja-6-ene (2) and a new *nor*-sesquiterpene, botocudorol (3). Spectral data were used in the structure determination. The ^1H and ^{13}C NMR chemical shifts have been assigned on the basis of APT, DEPT, ^1H x ^1H -HOMOCOSY and ^1H x ^{13}C -HETECOSY experiments.

Key words: *Sparattanthelium botocudorum*; *Hernandiaceae*; Flavonol; sesquiterpenes.

Introduction

Sparattanthelium botocudorum Mart, family Hernandiaceae, is a shrub widely distributed in the coastal region of Paraíba State and is commonly known as "canela-brava". A survey of the literature showed that alkaloids have been reported from *Sparattanthelium uncigerum*, the only species of the genus studied so far¹. In this paper we wish to describe the isolation and characterization of two known compounds, a flavonol (1) and a sesquiterpene (2), along with a new bisabolene type *nor*-sesquiterpenoid, botocudorol (3) from *Sparattanthelium botocudorum*.

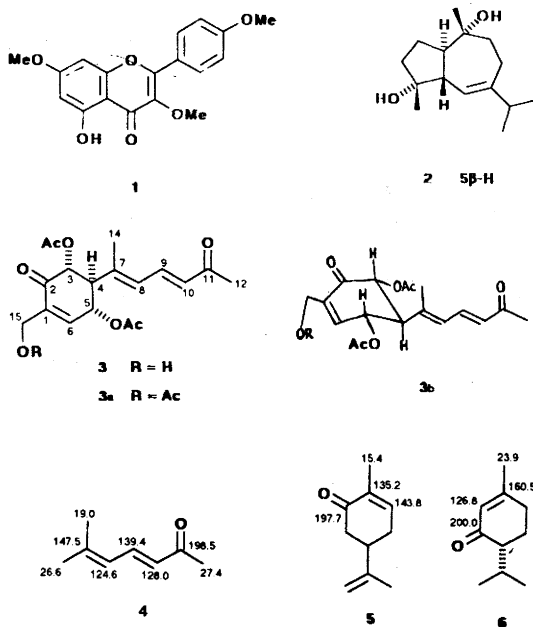
Experimental

General experimental procedures. Melting points were measured using a Kofler block and are uncorrected. ^1H (80 and 200 MHz) and ^{13}C (20 and 50.3 MHz) NMR spectra were obtained in CDCl_3 solution, on either a Varian FT-80A or a Bruker AC-200 equipment. TMS was used as an internal standard and the chemical shifts are given in δ values (ppm). EIMS were determined with a GC/MS system and relative intensities of peaks are reported with reference to the most intense peak higher than m/z 100. IR spectra

were recorded on a Perkin-Elmer 467 spectrometer. For TLC, Kieselgel 60 F₂₅₄ was used. Commercial Merck Si gel (60-230 mesh ASTM) was used for CC.

Plant material. The leaves of *Sparattanthelium botocudorum* were collected in the proximity of the city of João Pessoa by Maria de Fátima Agra, Universidade Federal da Paraíba, João Pessoa. A voucher specimen (n° 5799) is deposited at the Herbarium of the Universidade Federal da Paraíba, João-Pessoa, PB, Brazil.

Extraction and isolation of constituents. The pulverized and air-dried plant material (11 kg) was percolated with 95% EtOH at room temperature, and the EtOH extract was concentrated *in vacuo* to furnish a residue (800 g). This residue was suspended in 10% AcOH and extracted with CHCl_3 . The CHCl_3 solution was concentrated to give the CHCl_3 -soluble fraction (48 g) which was chromatographed on Si gel (700 g) column to give the combined fractions 15/16, 46/62 and 78/85, eluted with C_6H_6 - CHCl_3 (8:2); C_6H_6 - CHCl_3 (1:1) and C_6H_6 - CHCl_3 (1:2), respectively. These three fractions were further purified by Si gel TLC to furnish 1 (22 mg), 2 (62 mg) and 3 (115 mg), respectively.



5-Hydroxy-3,4',7-trimethoxyflavone (1). MP 143-145°C (benzene) (lit², mp 142-144°C). Spectral data (UV, IR ¹H and ¹³C NMR and EIMS) are in agreement with the reported values.^{2,3}

4α-10α-Dihydroxy-5β-H-guja-6-ene (2). Viscous oil. IR, ¹H NMR and EIMS data are consistent with those recorded in the literature;⁴ ¹³C NMR (50.3 MHz, CDCl₃).

TMS) δ149.6 (C-7), 121.3 (C-6), 80.2 (C-4), 75.3 (C-10), 50.6 (C-5), 50.3 (C-1), 42.7 (C-3), 40.4 (C-9), 37.3 (C-11), 25.1 (C-8), 22.5 (C-15), 21.5 (C-2), 21.4 (C-14), 21.3 (C-12), 21.2 (C-13).

Botocudorol (3). Viscous oil; ν(KBr,max) cm⁻¹: 3400, 1735, 1725, 1670, 1645, 1610, 1570, 1360, 1220, 1023; λ(MeOH,max, ε)nm: 227 (14.000), 287 (21.700); ¹H and ¹³C NMR: see Table 1; EIMS *m/z* (rel.int.) 350 (M⁺, absent), 332(1), 290(2), 272(1), 230(17), 215(5), 212(4), 202(8), 201(6), 187(22), 174(6), 159(12), 109(98), 43(100).

Acetate of 3 (3a). Compound 3 (10 mg) was dissolved in pyridine (1 ml) and Ac₂O (2 ml) was added, and the solution was allowed to stand for 24 h at room temperature. The usual workup afforded 3a (7 mg), viscous oil; ¹H and ¹³C NMR: see Table 1.

Results and discussion

The known compounds (1 and 2) were identified by comparison of spectral properties with literature data, as well as by ¹³C NMR spectra (see experimental). 5-Hydroxy-3,4',7-trimethoxyflavone (1) has been found in *Cheilanthes longissima*², *Cheilanthes farinosa*⁵, *Aframamum giganteum*⁶, *Sideritis dasygnaphala*⁷, *Sideritis bolleana*⁷, *Aesculus turbinata*⁸, *Ostrya virginiana*⁹ and *Betula nigra*⁹. 4α-10α-Dihydroxy-5β-H-guja-6-ene was also isolated earlier from *Silphium pefoliatum*⁴ and *Silphium terebinthinaceum*⁴.

The molecular formula C₁₄H₂₂O₇ for the *nor*-sesquiterpenoid botocudorol (3) was determined by a combination of low resolutions MS, and ¹H and ¹³C NMR (broadband decoupled, APT and DEPT), including 2D homonuclear (¹H x ¹H-HOMOCOSY) and heteronuclear (¹H x ¹³C-

Table 1. ¹H (200 MHz) and ¹³C (50.3 MHz) NMR data of natural product 3 and its acetate derivative 3a, run in CDCl₃ solution and TMS as internal standard.^a

C	δC	δH	δC	δH
1	137.8	-	137.7	-
2	192.1	-	192.1	-
3	72.5	5.46 (<i>d</i> , 13.1)	72.5	5.59 (<i>d</i> , 13.2)
4	56.7	3.15 (<i>dd</i> , 10.1, 13.1)	56.7	3.17 (<i>dd</i> , 10.8, 13.2)
5	68.1	5.88 (<i>d</i> , 10.1)	68.1	5.88 (<i>d</i> , 10.8)
6	142.2	6.77 (<i>br s</i>)	142.9	6.75 (<i>br s</i>)
7	142.6	-	142.8	-
8	129.4	6.07 (<i>d</i> , 11.3)	129.3	6.09 (<i>d</i> , 11.0)
9	137.7	7.34 (<i>dd</i> , 15.3, 11.3)	137.6	7.34 (<i>dd</i> , 15.3, 11.0)
10	131.0	6.13 (<i>d</i> , 15.3)	130.1	6.15 (<i>d</i> , 15.3)
11	198.8	-	198.8	-
12	27.6	2.28 (<i>s</i>)	27.5	2.29 (<i>s</i>)
13	-	-	-	-
14	13.4	1.87 (<i>br s</i>)	13.4	1.89 (<i>br s</i>)
15	59.4	4.30 (<i>br s</i>)	59.3	4.78 (<i>br s</i>)
OH	-	-	-	-
OAc	169.9 20.5	2.10 (<i>s</i>)	169.9 20.5	2.12 (<i>s</i>)
OAc	169.8 20.4	2.02 (<i>s</i>)	169.8 20.3	2.05 (<i>s</i>)
OAc	-	-	169.8 20.3	2.10 (<i>s</i>)

^a Chemical shifts are given downfield from TMS. Values of coupling constants (*J*) in Hz are reported in brackets. Assignments were made with the aid of the APT, DEPT and 2D-shift-correlated homonuclear (¹H x ¹H-HOMOCOSY) and heteronuclear [¹H x ¹³C-HETECOSY, optimized for one bond (¹JCH)].

HETECOSY) correlations¹⁰. In addition to the presence of two acetoxyl groups in the ¹H [δ 2.10 (s, 3H) and 2.02 (s, 3H) and ¹³C [δ 169.9 (s, COO), 169.8 (s, COO), 20.5 (q, CH₃) and 20.4 (q, CH₃)] NMR spectra, the remaining signals could be assigned to (C=O)₂, (C=CH)₂, (CH=CH), (CH₂OR), (CH-OR)₂, (CH) and (CH₃)₂.

The IR spectrum of compound 3 exhibited absorption bands at 3400 (OH), 1735 (OCOCH₃), 1725 (OCOCH₃), 1670 (α, β-unsaturated ketone), 1645 (α, β, γ, δ-unsaturated ketone), 1610 (C=C) and 1570 (C=C). The ¹H NMR spectrum revealed signals at δ 6.13 (d J=15,34 Hz, H-10)m 7,34 (dd, J=15.3 and J=11.3 Hz, H-9), 6.07 (d, J=11.3 Hz, H-8), 1.87 (br s, CH₃-7) and 2.28 (s, CH₃-11) which are characteristic of the 7-methyl-7,9-dien-11-one system [λ(MeOH,max)287 (ε 21.700)nm; calculated value: 215 + 30 + (2 x 18) = 281 nm]. The coupling constant (J=15.3 Hz) corresponding to the spin-spin interaction of H-9 with H-10 indicated a *trans* relationship between these two protons. The presence of this α, β, γ, δ-unsaturated methyl ketone system was confirmed by selective double resonance and ¹H x ¹H-HOMOCOSY experiments and by ¹³C NMR spectra (broadband decoupled, APT and DEPT) which showed signals at δ 198.8 (s, C-11), 142.6 (s, C-7), 137.7 (d, CH-9), 131.0 (d, CH-8), 129.4 (d, CH-10), 27.6 (q, CH₃-10 and 13.4 (q, CH₃-7). The assignments were made with the aid of the 2D-shift-correlated heteronuclear (¹H x ¹³C-HETECOSY) and comparison with model compounds, 6-methylhepta-3,5-dien-2-one(4)¹¹, 5 and 6¹². The α,β-unsaturated tetrasubstituted cyclohexenone ring [λ(MeOH,max)227, (14.000)nm] was diagnosed by the chemical shifts and multiplicities of signals of the ¹H [δ 5.46 (d, J=13.1 Hz, H-3, 3.15 (dd, J=10.1 and 13.1 Hz, H-4), 5.88 (d, J=10.1 Hz, H-5) and 6.77 (br s, H-6)] and ¹³C [d 137.8 (s, C-1), 192.1 (s, C-2), 72.5 (d, CH-3), 56.7 (d, CH-4), 68.1 (d, CH-5) and 142.2 (d, CH-6)] NMR. The existence of a primary allylic alcohol was recognized by a broad singlet at δ 4.30 which resulted in a 0.48 ppm paramagnetic shift upon acetylation(3a). The multiplicities of the signals corresponding to methine sp³ carbon atoms and the coupling constant values were used for the assignment of an axial position for H-3 (J=13.1 Hz), H-4 (J=10.1 and J=13.1 Hz) and H-5 (J=10.1 Hz). Consequently, the two acetoxyl groups and the methyl-7,9-dien-10-one system are placed at equatorial positions (3). These assignments were also supported by NMR techniques as described above (Table 1). Finally, the *E*-configuration for the double bond between C-7 and C-8 was indicated by upfield chemical shifts of C-14 (d 13.4) and C-9 (δ 137.7), showing an expected γ-effect. The considerable upfield shift of C-14 (13.4 ppm) in comparison with that observed for the analogous methyl group of the model compound 4 (19.0 ppm) is consistent with an additional effect of C-3 and C-5. This deduction was confirmed by NOE difference spectra¹⁰ of 3a performed with irradiation on methyl signal at δ 1.87 (br s, CH₃-7) which revealed H-3 (δ 5.49), H-5 (δ 5.88) and H-9 (δ 7.34) signal enhancements of 3.7, 3.5 and 3.4 %, respectively. The absence of an NOE at the H-4 during this experience shows that the C-7 is in a *trans* relationship with this proton. Thus the complete assignment of the relative configuration as shown in 3b was deduced for the new *nor*-sesquiterpenoid isolated from *Sparattanthelium botocudorum*. This *rel* - 3α, 5α,-

diacetoxyl-15-hydroxy-13-*nor*-4α-H-bisabola-6, 7*E*, 9*E*-trien-2, 11-di-one was named botocudorol.

References

1. M. C. Chalandre, H. Jacquemin and J. Bruneton. *J. Nat. Prod.* **48**, 333 (1985).
2. R. Sunder, K. N. N. Ayengar and S. Rangaswami, *Phytochemistry* **13**, 1610 (1974).
3. B. Voirin, *Phytochemistry* **22**, 2107 (1983).
4. F. Bohlmann and J. Jakupovic, *Phytochemistry* **18**, 1987 (1979).
5. H. Erdtmann, L. Novotny and M. Romani, *Tetrahedron* **22** (Suppl. 8) 1, 71 (1966).
6. G. Vidari, P.V. Finzi and M. Bernardi, *Phytochemistry* **10**, 3335 (1971).
7. A. G. Gonzales, B. M. Fraga, M. G. Hernandez, F. Larruga, J. G. Luis and A. G. Ravelo, *Lloydia* **41**, 279 (1978).
8. E. Wollenweber, *Z. Pflanzenphysiol* **73**, 277 (1974).
9. E. Vollenweber, *Biochem. Syst. Ecol.* **3**, 47 (1975).
10. A. E. Derome, *Modern NMR Techniques for Chemistry Research*, Pergamon Press, Oxford (1988).
11. E. Breitmaier and W. Voelter, *Carbon-13 NMR Spectroscopy: High-Resolution Methods and Applications in Organic Chemistry and Biochemistry* (3rd Edition), VCH, Weinheim (1987), p. 217 (Table 4.28).
12. F. W. Wehrli and T. Nishida, *Prog. Chem. Org. Nat. Prod.* **36**, 1(1976).