

New *nor*-Cucurbitacin Glucosides from *Wilbrandia* Species

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Duas novas *nor*-cucurbitacinas glicosídicas (**3** e **4**) foram isoladas das raízes de um espécime de *Wilbrandia* sp e suas estruturas foram elucidadas com base na análise de espectros de RMN uni- (1D) e bidimensionais (2D), em comparação com os dados das cucurbitacinas **1** e **2** isoladas da mesma espécie.

Two new *nor*-cucurbitacin glucosides (**3** and **4**) were isolated from the roots of a specimen of *Wilbrandia* sp. Their structures were elucidated by 1D and 2D NMR analysis, and comparison with the cucurbitacins **1** and **2** earlier isolated from the same source.

Key words: *Wilbrandia* sp; roots; *Cucurbitaceae*; *nor*-cucurbitacin glucosides; spectral data.

Introduction

We have previously reported¹ the isolation of two *nor*-cucurbitacin glucosides (**1** and **2**) from *Wilbrandia* species which were converted into their peracetyl derivatives **1a** and **2a** for separation and identification. In view of the medicinal potential of these species², we have now re-examined a larger amount of *Wilbrandia* sp extract that afforded again **1** and **2** in a mixture with two new *nor*-cucurbitacin glucosides **3** and **4**. The structure of compound **4** was elucidated on the basis of spectral data of its peracetyl derivative **4a**. 1D and 2D NMR analyses were performed and the new structures were correlated with the published NMR spectra of similar compounds.^{3,4}

Results and Discussion

In this paper we describe the isolation and identification of two new triterpenes **3** and **4** in mixtures with **1** and **2**, respectively, from the roots of a specimen of *Wilbrandia* sp.

The identification of cucurbitacin glucoside **1**, in the mixture of **1** and **3**, as 6,7-dehydrofelicordin A glucoside, a glucoside previously isolated from this plant,¹ allowed to recognize the presence of the additional signals in the ¹H (Table 1) and ¹³C NMR (Table 2) spectra consistent with the modification of the double bond localized between carbon atoms C-23 [**1**: δ_H 6.67 (d, J=15.1 Hz); δ_C 122.53 (d)] and C-24 [**1**: δ_H 7.06 (d, J=15.1 Hz); δ_C 151.61 (d)] to the corresponding 23,24-dihydroderivative (**3**: δ_H 2.7-2.5 (m, 2H-23), 1.4-1.1 (m, 2H-24); δ_C 32.78 (t, C-23) and 35.73 (t, C-24)].

Table 1. ^1H NMR (200 MHz) data of compounds **1** and **3** ($\text{CD}_3\text{OD} + \text{CDCl}_3$) and peracetyl derivatives **2a** and **4a** (CDCl_3), TMS as internal standard. The chemical shifts are expressed in δ (ppm) and the coupling constants (J) in Hz (in parenthesis).

H	1	1 + 3	3	2a	2a + 4a	4a
1		6.48(s)			6.45(s)	
6		6.79(d,10.1)			6.80(d,10.0)	
7		5.72(dd,10.1,5.8)			5.78(dd,10.0,6.4)	
8		2.46(d,5.8)			2.64(d,6.4)	
12		3.0-2.8			2.89(d,16.4)	
		2.7-2.5			2.77(d,16.4)	
15		2.0-1.8			a	
		1.3-1.1			a	
16		4.5-4.3			5.50(t,7.0)	
17		2.37(d,6.9)			2.57(d,7.1)	
18	0.88(s)		0.86(s)		0.91(s)	
19		1.29(s)			1.29(s)	
21		1.45(s)		1.47(s)		1.32(s)
22	-	-	-	-		5.00(d,7.1)
23	6.67(d,15.1)		2.7-2.5			5.62(dd,14.3,7.1)
24	7.06(d,15.1)		1.4-1.1			5.86(d,14.3)
26		1.34(s)			1.25(s)b	
27		1.31(s)			1.23(s)b	
28		2.14(s)			2.27(s)	
30		1.14(s)			1.09(s)	
1'		4.54(d,7.2)			4.80(d,7.1)	
2'		3.7-3.3			5.3-5.1	
3'		3.7-3.3			5.3-5.1	
4'		3.7-3.3			5.3-5.1	
5'		3.7-3.3			3.87(m)	
6'		3.90(dd,12.7,2.3)			4.30(dd,11.0,3.7)	
		3.76(dd,12.7,3.7)			4.23(dd,11.0,3.2)	
OAc	1.91(s)		1.85(s)		-	
					2.17(s)	
					2.13(s)	
					2.09(s)	
					2.07(s)	
					2.05(s)	
					2.04(s, 2x)	
					2.03(s)	
					1.94(s)	

^aOverlapped with other signals.

This modification was confirmed by the downfield shift ($\Delta\delta_{\text{C}}=215.52-204.39=11.13$ ppm) observed in the C-22 signal (δ 204.39) of **1** compared with the chemical shift of this carbon atom (δ_{C} 215.52) in **3**. As anticipated, this chemical shift alteration was also confirmed by reduction [**3**: δ_{C} 32.78 (t, C-23), 35.73 (t, C-24)] of the double bond [**1**: δ_{C} 122.53 (d, C-23), 151.61 (d, C-24)] of the conjugated carbonyl system. The remaining data obtained from the ^1H (Table 1) and ^{13}C NMR (Table 2) spectra was considered practically identical, since the less significant modifications observed in the chemical shifts of C-21 and C-25 were also used as additional support for the deduction described above. Thus, all the spectral data (Tables 1 and 2) of the new *nor*-cucurbitacin glucoside present in the mixture with **1** allowed its characterization as 23,24-dihydro-6,7-dehydrofevicordin A glucoside (**3**).

The mixture containing the *nor*-cucurbitacins **2** and **4** was transformed to peracetyl derivatives **2a** and **4a** by the usual treatment with acetic anhydride-pyridine. The analysis of the ^1H (Table 1) and ^{13}C NMR spectra (Table 2) revealed that the two peracetyl derivatives differ significantly only in the chemical shifts of the side-chain carbons at C-17. The remaining signals of ^1H and ^{13}C corresponding to this side-chain, after assignment of the absorptions belonging to **2a**, were assigned to the isomer **4a**, containing one hydroxy group at C-22 [δ_{H} 5.00 (d, $J=7.1$ Hz, H-22); δ_{C} 77.20 (d, C-22)] and one E double bond ($J=14.3$ Hz) localized between C-23 and C-24 [δ_{H} 5.62 (dd, $J=14.3$ and 7.1 Hz, H-23) and 5.86 (d, $J=14.3$ Hz, H-24); δ_{C} 120.20 (d, C-23) and 144.67 (d, C-24)] (Tables 1 and 2). The assignments of the signals of these protons were ascertained by the cross peaks in the homonuclear shift correlated 2D NMR ($^1\text{H}\times^1\text{H}$ -COSY) spec-

Table 2. ^{13}C NMR (50.3 MHz) data of compounds **1** and **3** ($\text{CD}_3\text{OD}+\text{CDCl}_3$) and peracetyl derivatives **2a** and **4a** (CDCl_3), TMS as internal standard. The chemical shifts are expressed in δ (ppm) and the multiplicity of the signals deduced by DEPT technique.

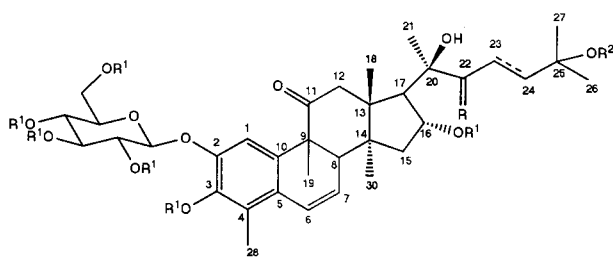
C	1	1+3	3	2a	2a + 4a	4a
1	112.53		112.40		108.38	
2	145.65		145.14		147.16	
3		129.68			137.00	
4		123.23			128.25	
5		128.17			126.19	
6		127.21			125.93	
7		126.09		124.66		124.45
8		46.12		45.98		46.00
9		50.41		50.51		50.74
10		129.68			135.04	
11	215.95		215.89 ^a		213.16	
12	52.05		51.89	51.12		51.41
13		48.85			47.63	
14		50.41			48.57	
15	44.74		44.64	41.23		41.89
16	71.77		72.13		74.64	
17	60.45		59.85	54.15		52.89
18		20.33			19.54	
19		18.05			17.08	
20	80.15		80.75	78.57		74.22
21	25.39		26.20	24.19		26.29
22	204.39		215.52 ^a	213.53		77.20
23	122.53		32.78	30.66		120.20
24	151.64		35.73	37.03		144.67
25	83.07		81.01		70.30	
26	26.63		26.74	29.34		29.83
27	26.63		26.29	29.34		29.83
28		11.28			11.77	
30	21.88		20.56		21.57-20.19	
1'		104.62			97.88	
2'		74.71			70.29	
3'		78.06			72.53	
4'		70.75			67.87	
5'		77.59			72.07	
6'		61.96			61.61	
OAc		20.54			21.38	
	172.45	20.32	171.89		21.10	
					20.59	
					20.51(2x)	
					20.19	
					171.11	
					170.90	
					170.15	
					169.43	
					169.36	
					168.52	

^a Signals interchangeable

trum, along with other spin-spin interaction of protons in the molecules of two components of the mixture of **2a** and **4a** (Tables 1 and 2). Thus, the structure of the peracetyl derivative was established as **4a** and, consequently, the structure of the additional new *nor*-cucurbitacin glucoside isolated from

Wilbrandia sp was defined as 22-dihydro-25-O-deacetyl-6,7-dehydrofevicordin A glucoside (**4**).

The assignments of the ^1H (Table 1) and ^{13}C (Table 2) chemical shifts of **3** and **4a** were based on the application of the usual chemical shift parameters, the observed multiplicity



	R	R ¹	R ²	
1	O	H	Ac	Δ ²³
1a	O	Ac	Ac	Δ ²³
2	O	H	H	23,24-dihydro
2a	O	Ac	H	23,24-dihydro
3	O	H	Ac	23,24-dihydro
3a	O	Ac	Ac	23,24-dihydro
4	H,OH	H	H	Δ ²³
4a	H,OAc	Ac	H	Δ ²³

of ¹H absorptions and ¹³C signals in the DEPT-¹³C NMR (number of attached protons for each carbon signal) spectra, the homonuclear 2D shift-correlated (¹Hx¹H-COSY) NMR spectrum⁵ (**4a**) and comparison with the corresponding values of the *nor*-cucurbitacin glucoside **1** and of the peracetyl derivative (**2a**) of **2**, isolated previously from the same *Wilbrandia* sp.¹

Experimental

General experimental procedures. M.p. were determined using a Kofler hot stage instrument and are not corrected. IR spectra were taken on a Perkin Elmer 257 spectrometer. ¹H and ¹³C NMR spectra were recorded in CD₃OD+CDCl₃ (**1** + **3**) and CDCl₃ (**2a** + **4a**), using TMS as internal standard employing a Bruker AC 200 (¹H: 200 MHz; ¹³C: 50.3 MHz) spectrometer.

Plant material. Roots of *Wilbrandia* sp. were collected at Senador Pompeu, Ceará State, Brazil. A voucher specimen (N° 13623) is deposited in the Herbarium Prisco Bezerra, Instituto de Biologia, Universidade Federal do Ceará, Fortaleza.

Extraction and isolation. Plant material (4 kg) was extracted with EtOH in a Soxhlet apparatus as previously described.¹ The residue (200 g), obtained after concentration of the extract under reduced pressure, was chromatographed on a silica gel (600 g) column. Elution with EtOAc-MeOH (4:1) furnished fraction A (18 g) and elution with MeOH gave fraction B (30 g). Fraction A (3 g) was rechromatographed on a silica gel (20 g) column and eluted with mixtures of increasing polarity of hexane and EtOAc. The fraction eluted with hexane-EtOAc (1:9) yielded the mixture of **1** and **3** (250 mg). The fraction B (4 g) was acetylated with Ac₂O (30 ml) and pyridine (10 ml) and chromatographed (4.5 g) on a silica gel column (30 g). The fraction eluted with hexane-EtOAc (1:3) yielded the mixture of **2a** and **4a** (150 mg).

Mixture of **1 and **3**.** Powder, m.p. 155-158 °C; IR (KBr), cm⁻¹: 3400 (OH), 1720-1680 (11,22-dioxo), 1620-1560 (aromatic); ¹H NMR: see Table 1; ¹³C NMR: see Table 2.

Mixture of **2a and **4a**.** Powder, m.p. 122-125 °C; IR (KBr), cm⁻¹: 3400 (OH), 1755 (esters), 1700 (11,22-dioxo), 1600, 1500 (aromatic); ¹H NMR: see Table 1; ¹³C NMR: see Table 2.

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